

## **EXHIBIT H**

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Northern RI Chapter 737 Trout Unlimited  
C/O Roland C. Gauvin  
2208 Mendon Rd.  
Cumberland, RI 02864  
September 22, 2008

US Environmental Protection Agency  
Clerk of the Board Environmental Appeals Board  
Colorado Building  
1341 G Street N.W., Suite 600  
Washington, DC20005

Re: NPDES Permit No. MA0102369

Dear Sirs:

The Northern RI Chapter 737Hearby contests the draft permit for the Upper Blackstone Water Pollution Abatement District 50 Route 20 Millbury, MA 01527.

In response to

Comment; #A7: Trout Unlimited commented that the permit should address concerns with aluminum toxicity.

Response #A7: We agree that aluminum toxicity is a potential concern. The final permit contains a monitoring requirement in order to obtain more information relative to the potential to violate receiving water criteria for aluminum. If the data indicates that there is a reasonable potential to violate receiving water criteria, future permit actions will include an aluminum limit.

In Comment #D2:EPA should utilize effluent data collected as part of the bioassay testing to determine whether reasonable potential exists for the UBWPAD facility to cause or contribute to water quality violations for additional pollutants. Since EPA does not enter pollutant data collected as part of the bioassay testing into ICIS, RIDEM was unable to evaluate reasonable potential for the following pollutants: Chromium, lead, nickel, and aluminum. At a minimum, based on typical lead levels seen in effluent from Rhode Island waste water treatment facilities, it appears that the UUBWPAD would have "reasonable potential" for lead and therefore would require lead limits. To ensure that bioassay pollutant monitoring data is readily available for review, RIDEM requests that EPA lists the pollutants monitored during the bioassay testing in Part1A1 of the permit.

Response#D2: We reviewed the bioassay reports from 2005 and 2006. The effluent chromium data are all below detection levels (detection levels ranged from 5-10 ug/l) and well below the applicable ambient criteria values in state standards. The effluent nickel data ranged from 5-20 ug/l which also is well below ambient criteria values. The effluent lead data are all below detection levels( detection levels ranged from 5-10 ug/l).

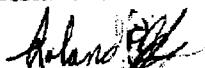
However the detection levels are higher than the ambient criteria values. Consequently we have

included a monthly lead monitoring requirement in the final permit, with a quantification level of 0.5 ug/l in order to be able to assess the need for a permit limit in future permit action. Effluent aluminum levels are of concern. Effluent values ranged from 70-240 ug/l. As indicated in response #A7, we have included a monthly monitoring requirement for aluminum in the final permit. A permit limit will be established if the data indicate a reasonable potential to exceed criteria.

We concur that requiring reporting of selected effluent data from bioassay testing on Discharge Monitoring Reports (in addition to submitting the information to EPA in a separate report) would make it easier to review these results. Copper, zinc, cadmium, aluminum, and lead are all required to be monitored more frequently than quarterly. Accordingly, for these metals, the final permit requires that the effluent results from the WET tests must be included in the required discharge monitoring reports. For nickel, a quarterly monitoring requirement has been included in the final permit in order that the effluent results for nickel from the WET tests are also included in the required discharge monitoring reports.

It is our contention that aluminum limits should be set at this time because in Response#A7 it states that limits will be set if data indicates that there is a potential to violate receiving water criteria. In Response#D2 it is stated that effluent levels of aluminum are of concern. Quote "Effluent values ranged from 70-240 eg/l. Because of these levels and documentation in exhibits A an B that substantiate that aluminum levels in this range are detrimental to the reproduction of salmonids. We implore the EPA to set discharge limits for aluminum and urge EPA to advocate for the use of technology that does not use aluminum oxide in the remediation of nitrogen discharge. We have invested much time money and effort in our project to bring fish ladders to the Blackstone and return anadromous fish to the river. Aluminum discharge at the current levels by the UBWPAD are unacceptable and discharge limits should be set at this time.

Roland C. Gauvin



For Northern RI chapter 737 Trout Unlimited

**LITIGATION ANALYSIS AND RECOMMENDATION FOR ADOPTION**

**AN INTEGRATED RIVER SYSTEMS APPROACH TO SALMON IDAHO**

**PLANNING AND MANAGEMENT**

**AN INTEGRATED RIVER SYSTEMS APPROACH TO SALMON IDAHO**

**PLANNING AND MANAGEMENT**

The following document is the result of a technical review and analysis of the salmon restoration plans developed by the State of Idaho and the U.S. Fish and Wildlife Service. It is intended to provide a critical assessment of the proposed actions and recommendations for improvement. The document is organized into several sections, each addressing a specific aspect of the restoration plan.

• Additional information regarding the proposed actions can be found in the following sections:

• **Background:** An introduction to the biology and ecology of salmon in Idaho, including habitat requirements, life history stages, and threats to survival.

• **Objectives:** The goals and objectives of the restoration plan, including the desired outcomes and timeframes.

• **Strategies:** The specific actions proposed to achieve the objectives, including habitat enhancement, hatchery programs, and other management measures.

• **Evaluation:** Methods for monitoring and evaluating the effectiveness of the restoration plan.

• **Conclusion:** Summary of findings and recommendations for improvement.

• Additional information regarding the proposed actions can be found in the following sections:

**1. Introduction**

Atlantic salmon formerly occurred in nearly every river system in the Gulf of Maine or the Hudson River, and annual returns are estimated to have been 500,000–1,000,000 fish. Reproducing stocks now exist in only seven rivers in Maine, and annual returns have declined to less than 30 in 1998 (USFWs, 1999). Harvest has been greatly reduced and the rivers are stocked with hatchery-produced fish; but populations have failed to increase. Atlantic salmon populations have been reduced by acidic deposition in Nova Scotia, Canada (Lacroix, 1980) and Norway (Hesthagen and Hansen, 1991); however, in previous investigations we were unable to demonstrate significant mortality of river resident life stages of Atlantic salmon in Maine due to acidity (Mills et al., 1998). Recently, Chapman et al. (1998) documented that juvenile salmon in the St. Croix River in New Brunswick, Canada, and the Kennebec River in Maine, USA, have suffered significant mortality due to acidification during their first year of life, which may be contributing to the

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**EXHIBIT**

**A**

in the river system. A comparison of the estimated mean water velocity at hatchery and release sites indicated that the water velocity at the hatchery was slightly higher than at the release site (Table 1).

### 2.3.2. Hatchery

The hatchery (Fig. 3) was located approximately 100 m upstream from the mouth of the creek and 1.5 km upstream from the confluence of the creek with the river. The hatchery facility consists of a concrete building with a floor area of 100 m<sup>2</sup> containing two separate rearing tanks. Each tank has a volume of 25 m<sup>3</sup> and a water depth of 1.2 m. The tanks are constructed of concrete and have a thickness of 0.3 m. The tanks are connected by a 2.5 cm diameter tube. The water temperature in the tanks is monitored daily and the water level is checked weekly. The water in the tanks is renewed weekly and the water is filtered through a sand filter. The water is then chlorinated (50 mg/L chlorine) and then added to each tank. Chlorine was added at 50 mg/L from bleach solution into a mixing chamber with a reaction time of 30 min. An unchlorinated stream sample, a year older than hatchery rearing or spawning site, individual稚鱼 (36 mm) were collected during early October (Edmunds et al., 1994) and were used to determine water values. Unchlorinated smolt or smolt from Pender River creek rearing tank ( $18.5 \pm 2.18$  g/m<sup>3</sup> mean weight  $\pm$  S.E. =  $67.07 \pm 11.78$  g/m<sup>3</sup> n=14) were obtained from the Gifford Pinchot National Forest Hatchery in Ellsworth, Alaska and 50 each were placed in separate tanks and acclimated to the hatchery tanks. Water determined to be equal to the water quality in the creek and hatchery tanks was added to each tank. The water was added to each tank until the water level was 1.2 m. The water temperature was 10°C and the pH was 7.0. The water in the tanks was renewed weekly. The water in the tanks was filtered and chlorinated weekly. The water was filtered through a sand filter and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank. The water was chlorinated weekly and then added to each tank.

Smolt migration behavior was monitored with the use of surgically implanted ultrasonic transmitters (4.5 cm x 2.0 mm; Vemco, 1996) and stationary receivers in the river system (Hume, 1991). Hatchery and wild smolt were released in groups of 3 to 5 on three occasions between 10 and 16 May and were tracked until 28 May.

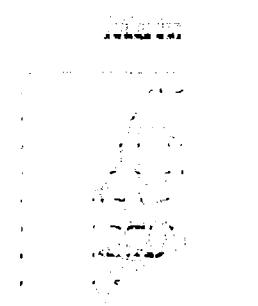


FIGURE 3  
Map of the river system showing the location of the hatchery and the river's course downstream.

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Leicestershire, UK

<sup>1</sup>See also Chapter 3.3.2.1 for a detailed description of the hatchery facility.

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water would cause  
the decrease in survival.

and 1970 (1970).  
The first 10 smolt were  
from hatchery rearing  
with additional 44  
smolt gradually in  
each 4 months to 1973.  
A physiology study  
which simultaneously  
monitored river which  
was Klamath and one  
smolt S.D.D. finger  
smolt added to either  
the Klamath or the  
Sacramento River at  
the time of release.  
After 10 days, the  
smolt were released  
into the Sacramento  
River. The smolt  
which were placed in  
Klamath were assumed  
to be smolts  
in smolt migration  
on 1 May. Three to  
May smolts were  
smolt within two months.  
ATPase activity  
was measured in the  
smolt at the  
time of release  
and again at the  
time of capture by  
Bart et al., 1981). All  
selected methods of

use of smolting in  
the Sacramento River  
are reported to  
be reliable (Bart  
et al., 1981).



Figure 2. Map showing location of study area.

All water physico-chemical properties data for standardization (USEPA, 1987), 15 km, 30 km upstream and downstream, were collected from the Sacramento River at the time of smolt migration. The 15 km upstream and downstream of the smolt release point was used to determine the influence of the Klamath River on the smolt movement. Differences between the two rivers were determined by the paired t-test (Sokal, 1974). The Wilcoxon Williams test was used to determine differences between Sacramento with respect to time of movement (Swanson, 1981). Mortality in each river section was estimated using the MARK computer program with the Cormack-Jolly-Seber model (White, 1990).

### 3. Results

No hatchery-reared smolts from any treatment died in RM or SW, but of the 24 h in SW one wild smolt died, one was immobile, and one was dead on arrival overnight. While smolts from RM, 15 km upstream, had a higher ATPase activity than smolts from RM, 30 km upstream, the difference was not significant (Table 1). The smolts from RM, 30 km upstream, had a significantly higher ATPase activity than smolts from RM, 15 km upstream, and smolts from SW (Table 1). No significant difference was found between smolts from RM, 15 km upstream, and SW (Table 1).

water, which may be due to the fact that the fish were acclimated to the river water prior to the experiment. The results of the present study are similar to those of other authors (Dobson et al., 1974; Dobson and

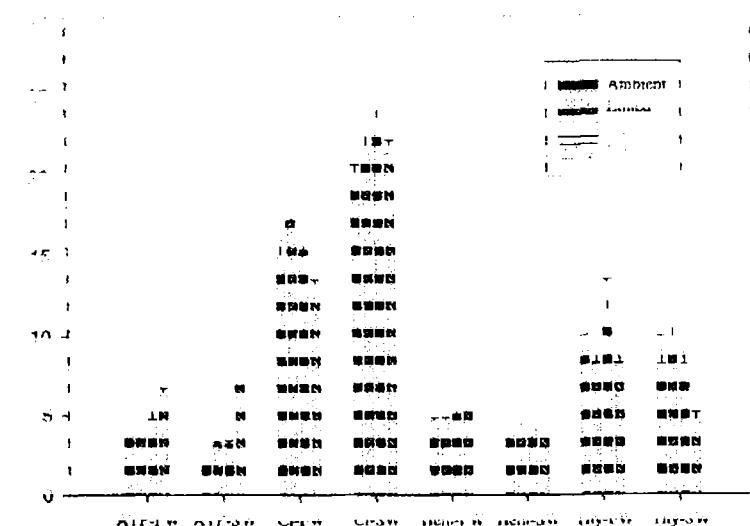


Fig. 2. Multiple regression analysis of the relationship between the following variables: ALB (mg/dl), ALPase (U/mg/mg), Cu (microg/l/l), Hem (hematocrit (%/10)), and Myoglobin (microg/l/l).

Table 1. Results of the multiple regression analysis of the relationship between the following variables: ALB (mg/dl), ALPase (U/mg/mg), Cu (microg/l/l), Hem (hematocrit (%/10)), and Myoglobin (microg/l/l) after 15 days of exposure to different concentrations of river water.

Treatment	Residence time in days				
	1	2	3	4	5
Control	1.00	1.00	1.00	1.00	1.00
1 mg/l	1.00	1.00	1.00	1.00	1.00
10 mg/l	1.00	1.00	1.00	1.00	1.00
100 mg/l	1.00	1.00	1.00	1.00	1.00
1000 mg/l	1.00	1.00	1.00	1.00	1.00

After 15 days of exposure to different concentrations of river water, the following results were obtained: the upper section of the river was characterized by relatively low concentrations of river water (1 mg/l), while the middle section (10 mg/l) and the lower section (100 mg/l) had relatively high concentrations of river water. The results of the present study are similar to those of other authors (Dobson et al., 1974; Dobson and

McDonald, 1975; Dobson and McDonald, 1976; Dobson et al., 1977). The results of the present study also show that the fish exposed to 100 mg/l of river water had a significantly higher mortality rate than those exposed to 10 mg/l and 1 mg/l of river water. The results of the present study also show that the fish exposed to 100 mg/l of river water had a significantly higher mortality rate than those exposed to 10 mg/l and 1 mg/l of river water.

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